

Prepared in cooperation with the U.S. Centers for Disease Control and Prevention and the Maine Center for Disease Control and Prevention

# Methods of Collection and Quality Assessment of Arsenic Data in Well-Water Supplies in Maine, 2001–2 and 2006–7

Data Series 1125

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U.S. Department of the Interior

**U.S. Geological Survey** 

Data Series 1125

# **U.S. Department of the Interior** DAVID BERNHARDT, Secretary

#### **U.S. Geological Survey**

James F. Reilly II, Director

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### **Contents**

Anstract		l
Introduc	tion	1
Pur	pose and Scope	1
Des	scription of Study Area	2
Methods	s of Data Collection	2
Me	asurement of Field Water-Quality Constituents	2
Sar	nple Collection for Well-Water Chemical Analyses Performed in the Field	2
Poi	nt-of-Entry and Point-of-Use Sample Collection for Laboratory Chemical Analysis of Well Water	2
Poi	nt-of-Entry and Point-of-Use Sample Collection for Laboratory Arsenic and Arsenic Speciation (Valence) Analysis	8
Ars	enic Sample Preparation and Preservation	8
Pre	paration of Field Anion-Exchange Chromatography Columns for Arsenic Speciation Analysis	8
Quality A	Assessment of Laboratory Arsenic Analysis	9
Referen	ces Cited	10
Figure	s	
1.	Map showing domestic-well sample locations in Maine, 2001–2 and 2006–7	3
2.	Boxplots showing distribution of unfiltered arsenic concentrations in paired samples analyzed by the Maine Department of Human Services Health and Environmental Testing Laboratory (HETL) and Underwriters Laboratories Inc. (UL), 2006–7	q
3.	Graph showing arsenic concentrations in paired samples analyzed by the Maine Department of Human Services Health and Environmental Testing Laboratory and Underwriters Laboratories Inc., 2006–7	
Tables		
1.	Method information, minimum reporting limits, and U.S. Geological Survey National Water Information System parameter codes for all constituents measured in water samples collected from domestic wells in Maine for laboratory analyses, 2001–2 and 2006–7	4
2.	Constituents, U.S. Geological Survey National Water Information System parameter codes, minimum reporting limits, and method information for all water samples collected from domestic wells in Maine for analyses performed in the field, 2001–2 and 2006–7	5
3.	Constituents, field parameters, arsenic speciation, and measuring entities for water samples collected from domestic wells in Maine, 2001–2 and 2006–7	7
4.	Representative efficiency of anion-exchange chromatography resin for the field separation of As(III) and As(V) for single arsenic species recovery	
5.	Representative efficiency of anion-exchange chromatography resin for the field separation of As(III) and As(V) for As(III)+As(V) mix recovery	

#### **Conversion Factors**

International System of Units to U.S. customary units

Multiply	Ву	To obtain
	Length	
micrometer (µm)	0.00003937	inch (in.)
centimeter (cm)	0.3937	inch (in.)
meter (m)	3.281	foot (ft)
kilometer (km)	0.6214	mile (mi)
	Volume	
cubic meter (m³)	6.290	barrel (petroleum, 1 barrel = 42 gal)
milliliter (mL)	0.03381	ounce, fluid (fl. oz)
	Flow rate	
liter per second (L/s)	15.85	gallon per minute (gal/min)
	Mass	
milligram (mg)	0.00003527	ounce, avoirdupois (oz)
kilogram (kg)	2.205	pound, avoirdupois (oz)

Temperature in degrees Celsius (°C) may be converted to degrees Fahrenheit (°F) as  $^{\circ}F = (1.8 \times ^{\circ}C) + 32$ .

#### **Datum**

Horizontal coordinate information is referenced to the North American Datum of 1983 (NAD 83).

# **Supplemental Information**

Specific conductance is given in microsiemens per centimeter at 25 degrees Celsius ( $\mu$ S/cm at 25 °C).

Concentrations of chemical constituents in water are given in milligrams per liter (mg/L) or micrograms per liter ( $\mu$ g/L).

Turbidity is reported in nephelometric turbidity units (NTU).

Concentrations of bacteria are reported in colony forming units per 100 milliliters (CFU/100 mL).

# **Abbreviations**

EPA U.S. Environmental Protection Agency

HETL Health and Environmental Testing Laboratory, Maine Department of Human Services

NWQL National Water Quality Laboratory

POE point of entry
POU point of use

QA/QC quality assurance and quality control

UL Underwriters Laboratories Inc.

USGS U.S. Geological Survey

# Methods of Collection and Quality Assessment of Arsenic Data in Well-Water Supplies in Maine, 2001–2 and 2006–7

By Charles W. Culbertson,<sup>1</sup> James M. Caldwell,<sup>1</sup> Luther F. Schalk,<sup>1</sup> Deana Manassaram,<sup>2</sup> Lorraine C. Backer,<sup>2</sup> and Andrew E. Smith<sup>3</sup>

#### **Abstract**

The U.S. Geological Survey, in cooperation with the U.S. Centers for Disease Control and Prevention and the Maine Center for Disease Control and Prevention, assessed the chemical characteristics and the occurrence, distribution, and oxidation state of inorganic arsenic in drinking water from selected domestic well-water supplies in Maine in 2001–2 and 2006–7.

The data collected provide support for evaluating arsenicremoval efficiencies of household water-purification systems and provide information to State and local officials that can be used in determining a water-treatment approach for the removal of arsenic from drinking water.

#### Introduction

Arsenic is toxic to humans and widespread in surface water and groundwater throughout the United States and world (National Research Council, 1999; Welch and others, 1999; Welch and others, 2000; Focazio and others, 2000; Ryker, 2001; Smedley and Kinniburgh, 2002; DeSimone and others, 2009). Arsenic in surface waters and groundwater aquifers has both anthropogenic and natural sources. Anthropogenic sources include current and historical industrial uses, as well as broad-spectrum uses in the lumber, agriculture, and livestock industries (National Research Council, 2001). Arsenic occurs naturally in the Earth's crust, ranking as the 20th most abundant element, with an average crustal abundance of approximately 2 milligrams per kilogram (0.0002 percent). It is common in weathered volcanic and marine sedimentary rocks containing iron and manganese oxides, in fossil fuels, and in a number of minerals, including arsenopyrite (FeAsS), orpiment (As<sub>2</sub>S<sub>3</sub>), scorodite (FeAsO<sub>4</sub>), and realgar (AsS) (Cullen and Reimer, 1989; Korte and Fernando, 1991; Smedley and Kinniburgh, 2002).

Arsenic in groundwater is primarily inorganic, either in the oxidized state as arsenate [As(V)], the form typical in oxic waters, or the reduced state as arsenite [As(III)], the form typical in anoxic waters (Cullen and Reimer, 1989; Smedley and Kinniburgh, 2002).

Roughly half of the population of Maine derives its drinking water from domestic wells, of which about 75 percent are drilled bedrock wells. Arsenic concentrations above the current U.S. Environmental Protection Agency (EPA) drinking-water standard of 10 micrograms per liter (µg/L or parts per billion) (U.S. Environmental Protection Agency, 2001) have been detected in water from drilled bedrock wells throughout Maine (Marvinney and others, 1994, 1995; Peters and others, 1999; Ayotte and others, 1999, 2003, 2006; Lipfert and others, 2006; Yang and others, 2009; Nielsen and others, 2010; Zheng and Ayotte, 2015; Flanagan and others, 2012). Effective removal of arsenic by household water-purification systems is constrained by the chemical form (for example, oxidation state) in which it occurs, as well as the prevailing chemical characteristics of the well water, including pH, and the presence of competing ions, such as phosphate (U.S. Environmental Protection Agency, 2001).

The U.S. Geological Survey (USGS), in cooperation with the U.S. Centers for Disease Control and Prevention and the Maine Center for Disease Control and Prevention, assessed the chemical characteristics and the occurrence, distribution, and oxidation state of inorganic arsenic in drinking water from selected domestic well-water supplies in Maine in 2001–2 and 2006–7.

The data collected provide support for evaluating arsenicremoval efficiencies of household water-purification systems and provide information to State and local officials that can be used in determining a water-treatment approach for the removal of arsenic from drinking water.

#### **Purpose and Scope**

This report describes methods used to collect arsenic and other water-chemistry data from domestic well water in support of studies evaluating arsenic removal efficiencies of household water purification systems. Data consist of water-chemistry results from samples collected over two time periods: 2001–2 and 2006–7 (also two samples collected

<sup>&</sup>lt;sup>1</sup>U.S. Geological Survey.

<sup>&</sup>lt;sup>2</sup>U.S. Centers for Disease Control and Prevention.

<sup>&</sup>lt;sup>3</sup>Maine Department of Health and Human Services.

in 2008), well characteristics (depth and type), and chemical characteristics of well water from approximately 120 selected domestic wells in known high-arsenic clusters throughout the State of Maine. All data are available in the USGS National Water Information System (NWIS) database, are searchable by USGS site number (U.S. Geological Survey, 2019), and are published in Culbertson and others (2020).

#### **Description of Study Area**

The study area includes most of the populated areas in the State of Maine apart from Aroostook County in northeastern Maine. The samples collected were from singlefamily dwellings relying on private well water as the primary drinking-water supply; additionally, these households had water-purification systems designed for the removal of arsenic installed either at the well-water point of entry (POE) or point of use (POU). The initial pilot study included 31 households sampled for arsenic and arsenic species in 2001–2, whereas the full-scale study included 120 households sampled in 2006–7. Households were recruited based on their locations in areas of Maine known to have high concentrations of arsenic  $(>10 \mu g/L)$  in groundwater (fig. 1).

#### **Methods of Data Collection**

Households with domestic wells included in this study were identified and prescreened prior to both sampling periods by project personnel associated with concurrent studies by the U.S. Centers for Disease Control and Prevention and the Maine Center for Disease Control and Prevention. To be considered for the study, households needed to have well water containing elevated (>10 µg/L) concentrations of arsenic and a previously installed water-purification system designed to remove arsenic; systems used reverse-osmosis, anionexchange, or adsorption technologies. Most homes tested had point-of-use reverse-osmosis systems installed. Well-water samples (pre-water-treatment system) were collected at the house POE, prior to the pressure tank and household watertreatment system(s), by using the existing domestic well-water pump. Samples collected after water treatment were collected at the kitchen (POU) faucet. A list of constituent names, USGS NWIS parameter codes, and minimum reporting limits for all samples collected for laboratory and field analyses are shown in tables 1 and 2. A summary of field measurements and analytical determinations, along with the entity or entities that analyzed or measured each, is presented in table 3.

#### **Measurement of Field Water-Quality Constituents**

Prior to collection of POE well water for arsenic and other chemical analyses, field measurements of temperature, specific conductance, pH, and dissolved oxygen were measured onsite using a multiparameter water-quality monitor immersed in a flow-through chamber under a steady water flow of approximately 1–1.5 liters per minute. Field measurements and water-chemistry results are published in Culbertson and others (2020). During the 2006–7 sample collection period, low dissolved oxygen readings (<1 mg/L) were confirmed by an alternate, colorimetric method (CHEMets colorimetric dissolved oxygen test kit, 0-1 part per million; CHEMetrics, Inc., Calverton, Va.). Samples for water chemistry determinations and arsenic analyses were collected after water-quality-monitor readings had stabilized (generally 0.25-0.75 hour). Except for specific conductance, field measurements of the other water-quality constituents in POU water samples were not made because of physical and chemical alterations imparted by the water-treatment systems. Specific conductance, a proxy for total dissolved solids, was measured in both prefiltration and postfiltration water, specifically to assess the performance efficiencies of reverse-osmosis water-treatment systems in removing arsenic (Fravel and Lindsey, 2014).

#### Sample Collection for Well-Water Chemical **Analyses Performed in the Field**

Upon obtaining stabilized readings of temperature, specific conductance, and pH, well-water samples for field chemical analyses were collected. Field measurements (or analyses) of POE well water included total iron, ferrous iron (FeII), total manganese, reactive phosphorus, sulfate, sulfide, and nitrate. These measurements were made immediately upon sample collection using standard colorimetric methods (American Public Health Association, 1998; Hach Company, 2000). Equipment blanks and sample replicates for quality-control (QC) purposes were collected and analyzed with the field samples. Additionally, during the 2006–7 sample collection period, replicate samples for some constituents were collected for chemical analysis by an alternate laboratory and method (Underwriters Laboratories Inc. [UL], South Bend, Indiana). Results of field chemical and quality-control analyses are published in Culbertson and others (2020).

#### Point-of-Entry and Point-of-Use Sample **Collection for Laboratory Chemical Analysis of** Well Water

According to study protocols for the 2006–7 study, two laboratories (Underwriters Laboratories Inc. [UL], South Bend, Indiana, and the Health and Environmental Testing Laboratory [HETL] of the Maine Department of Human Services<sup>4</sup> in Augusta, Maine) were used to measure arsenic concentrations. The laboratory results from these two facilities are published here as a means of comparison of the analytical

<sup>&</sup>lt;sup>4</sup>Since July 2004, the Maine Department of Health and Human Services.

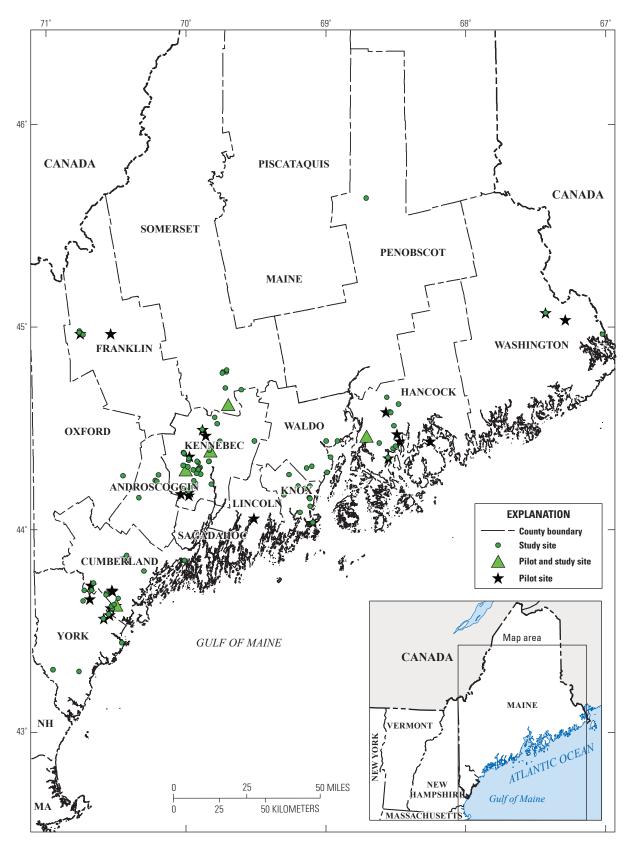


Figure 1. Domestic-well sample locations in Maine, 2001–2 and 2006–7.

4

Table 1. Method information, minimum reporting limits, and U.S. Geological Survey National Water Information System parameter codes for all constituents measured in water samples collected from domestic wells in Maine for laboratory analyses, 2001–2 and 2006–7.

methods, or U.S. Geological Survey (USGS) National Water Quality Laboratory analytical methods; MCL, maximum contaminant level; SMCL, secondary maximum contaminant level; MRL, minimum report-Environmental Testing Laboratory; NWQL, National Water Quality Laboratory; As, arsenic; CFU/100 mL, colony forming units (colonies) per 100 milliliters; N, nitrogen; NTU, nephelometric turbidity units] ing limit; N/A, not applicable; mg/L, milligram per liter; CaCO<sub>3</sub>, calcium carbonate; UL, Underwriters Laboratories Inc.; µg/L, microgram per liter; HETL, Maine Department of Human Services Health and Samples were collected for laboratory measurements before (point-of-entry, POE) and after (point-of-use, POU) household water-treatment systems. CASRN, Chemical Abstracts Service Registry Number; method reference, indicates the method code number associated with U.S. Environmental Protection Agency analytical methods, Standard Methods for the Examination of Water and Wastewater analytical

Constituent	CASRN1	Method reference	MCL/ SMCL	Unit	MRL	USGS param-	USGS method	Analyzing entity	Analyte description
Alkalinity, total	N/A	2320 B <sup>2</sup>	N/A	mg/L as CaCO <sub>3</sub>	10	29803	090LL	nr	Alkalinity, water, filtered, Gran titration, laboratory, milligrams per liter as calcium carbonate.
Arsenic	7440-38-2	200.83	10	μg/L	5	01002	PM100	NT	Arsenic, water, unfiltered, micrograms per liter.
Arsenic	7440-38-2	200.83	10	μg/L	0.5	01002	PLM99	HETL	Arsenic, water, unfiltered, micrograms per liter.
Arsenic	7440-38-2	$\mathrm{GF096^4}$	10	ng/L	1.9	01002	GF096	USGS- NWQL	Arsenic, water, unfiltered, micrograms per liter.
Arsenic	7440-38-2	200.83	10	μg/L	0.5	01000	PLM98	HETL	Arsenic, water, filtered, micrograms per liter.
Arsenite	15502-74-6	200.83	10	μg/L as As	0.5	62452	PLM98	HETL	Arsenite, water, filtered, micrograms per liter as arsenic.
Arsenate	15584-04-0	200.83	10	μg/L as As	0.5	62453	PLM98	HETL	Arsenate, water, filtered, micrograms per liter as arsenic.
Fecal coliform	N/A	$9222~\mathrm{D}^2$	-	CFU/100 mL	_	31616	BAC93	nr	Fecal coliform, M-FC MF (0.45 micrometer) method, water, colony forming units per 100 milliliters.
Fluoride	16984-48-8 300.03	300.03	4	mg/L	0.1	00951	IC075	nr	Fluoride, water, unfiltered, milligrams per liter.
Hardness, total	N/A	2340 B <sup>2</sup>	N/A	mg/L as CaCO <sub>3</sub>	10	60600	660LL	UL	Hardness, water, unfiltered, measured, milligrams per liter as calcium carbonate.
Iron, total	7439-89-6	200.73	0.3	mg/L	0.02	74010	PLA44	nr	Iron, water, unfiltered, milligrams per liter.
Lead	7439-92-1	200.83	15	μg/L	5	52298	PM100	nr	Lead, water, unfiltered, micrograms per liter.
Manganese	7439-96-5	200.83	50	µg/L	5	71883	PM100	nr	Manganese, water, unfiltered, micrograms per liter.
Nitrate	14797-55-8	353.23	10	mg/L as N		00630	CDR25	nr	Nitrate plus nitrite, water, unfiltered, milligrams per liter as nitrogen.
Nitrite	14797-65-0	353.23	_	mg/L as N	0.1	00615	CDR25	nr	Nitrite, water, unfiltered, milligrams per liter as nitrogen.
Hd	N/A	150.13	6.5 - 8.5	pH units	N/A	00403	EL022	nr	pH, water, unfiltered, laboratory, standard units.
Sodium	7440-23-5	200.73	N/A	mg/L	0.1	52290	PLA44	nr	Sodium, water, unfiltered, milligrams per liter.
Sulfate	14808-79-8	300.03	250	mg/L	10	00946	IC075	nr	Sulfate, water, unfiltered, milligrams per liter.
Turbidity	N/A	$180.1  \mathrm{LL}^3$	N/A	NTU	0.1	82079	<b>TBD05</b>	nr	Turbidity, water, unfiltered, laboratory, nephelometric turbidity units.
Uranium	7440-61-1	200.83	30	µg/L	5	63032	PM100	UL	Uranium, water, unfiltered, micrograms per liter.

<sup>1</sup> This report contains CAS Registry Numbers, which is a Registered Trademark of the American Chemical Society. CAS recommends the verification of the CASRNs through CAS Client Services.

<sup>&</sup>lt;sup>2</sup>Analytical method code from "Standard Methods for the Examination of Water and Wastewater" (American Public Health Association, 1998)

<sup>&</sup>lt;sup>3</sup> Analytical method code from the U.S. Environmental Protection Agency.

<sup>&</sup>lt;sup>4</sup>Analytical method code from the USGS National Water Quality Laboratory.

Constituents, U.S. Geological Survey National Water Information System parameter codes, minimum reporting limits, and method information for all water samples collected from domestic wells in Maine for analyses performed in the field, 2001–2 and 2006–7. Table 2.

[USGS, U.S. Geological Survey; MRL, minimum reporting limit; mg/L, milligram per liter; CASRN, Chemical Abstracts Service Registry Number; N, nitrogen; wu, water, unfiltered; NEWSC, New England Water Science Center; PO4, phosphate; °C, degree Celsius; NA, not applicable; µS/cm at 25 °C, microsiemens per centimeter at 25 degrees Celsius; ppm, part per million; % saturation, percent saturation; std units, standard units; gal/min, gallon per minute]

Constituent	USGS parameter code	Units	MRL (mg/L)	CASRN¹	Analytical method number reference <sup>2</sup>	USGS method code	USGS method name	Method description
					Field chemistry parameters	try parame	iters	
Nitrate, water, unfiltered, field, milligrams per liter as nitrogen	99124	mg/L as N	0.1	14797-55-8	8039	CDR23	Nitrate, wu, field, colorimetry, Hach test kit (NEWSC)	Field determination of nitrate in unfiltered water by cadmium reduction colorimetry method with Hach Test Kit.
Sulfate, water, unfiltered, field, milligrams per liter	99127	mg/L		14808-79-8	8051	CL203	Sulfate, wu, field, colorimetry, Hach test kit (NEWSC)	Field determination of sulfate in unfiltered water with Hach Test Kit, SulfaVer 4 method.
Orthophosphate, water, unfiltered, milligrams per liter as PO4	99126	mg/L	0.01	7723-14-0	8048	ASC20	Phosphate (orthophosphate), wu, field, colorimetry, Hach test kit (NEWSC)	Field determination of phosphate in unfiltered water with Hach Test Kit, PhosVer 3 (ascorbic acid) method.
Iron(II), water, unfiltered, field, milligrams per liter	99128	mg/L	0.01	15438-31-0	8146	PHEN5	Iron(II), wu, field, Hach colorimetry	Iron(II) in unfiltered water determined in the field by the 1,10-phenanthroline method with portable colorimeter using Hach AccuVac ampules. Range up to 3.00 mg/L.
Iron, water, unfiltered, field, 99138 milligrams per liter		mg/L	0.01	7439-89-6	8008	CL204	Iron, total, wu, field, colorimetry, Hach test kit (NEWSC)	Field determination of total iron in unfiltered water with Hach Test Kit, FerroVer method. Range up to 3.00 mg/L.
Manganese, water, unfiltered, field, milligrams per liter	53570	mg/L	0.1	7439-96-5	8034	CL205	Manganese, wu, field, colorimetry, Hach test kit (NEWSC)	Field determination of manganese in unfiltered water with Hach Test Kit, Periodate Oxidation method. Range up to 20.00 mg/L.
Sulfide, water, unfiltered, field, milligrams per liter	99119	mg/L	0.001	18496-25-8	8131	MB002	Sulfide, field, by Hach methylene blue colorimetric method test kit	Sulfide, field, by Hach methylene blue colorimetric method test kit.

Table 2. Constituents, U.S. Geological Survey National Water Information System parameter codes, minimum reporting limits, and method information for all water samples collected from domestic wells in Maine for analyses performed in the field, 2001–2 and 2006–7.—Continued

[USGS, U.S. Geological Survey; MRL, minimum reporting limit; mg/L, milligram per liter; CASRN, Chemical Abstracts Service Registry Number; N, nitrogen; wu, water, unfiltered; NEWSC, New England Water Science Center; PO4, phosphate; °C, degree Celsius; NA, not applicable; µS/cm at 25 °C, microsiemens per centimeter at 25 degrees Celsius; ppm, part per million; % saturation, percent saturation; std units, standard units; gal/min, gallon per minute]

Constituent	USGS parameter code	Units	MRL (mg/L)	CASRN1	Analytical method number reference <sup>2</sup>	USGS method code	USGS method name	Method description
					Field water-quality parameters	Jality param	eters	
Temperature, water, degrees Celsius	000010	O.	NA	NA	NA	THM01	Temperature, water, wu, field, thermistor, multiparameter sonde (NEWSC)	Field determination of water temperature (° C) in unfiltered water with multiparameter sonde.
Specific conductance, water, unfiltered, micro- siemens per centimeter at 25 degrees Celsius	96000	μS/cm at 25 °C	Z A	NA	NA	SC001	Specific conductance, wu, field, electrometry, multipa- rameter sonde (NEWSC)	Field determination of specific conductance in unfiltered water by electrometry with multiparameter sonde.
Dissolved oxygen, water, unfiltered, milligrams per liter	00300	mg/L	0.01	7782-44-7	NA	LUMIN	Dissolved oxygen, wu, field, luminescence sensor, multi- parameter sonde (NEWSC)	Field determination of dissolved oxygen in unfiltered water by luminescence-based sensor with multiparameter sonde.
Dissolved oxygen, water, unfiltered, milligrams per liter	00300	mdd	0.01	7782-44-7	NA	RHODA	Dissolved oxygen, wu, field, CHEMetrics 0–1 ppm kit (NEWSC)	Field determination of dissolved oxygen in unfiltered water with CHEMetrics indigo carmine test kit, 0–1 ppm (mg/L).
Dissolved oxygen, water, unfiltered, percent of saturation	00301	% satura- tion	0.1	7782-44-7	NA	CAL35	Dissolved oxygen solubility, barometric pressure, temperature, calculation, multiparameter sonde (NEWSC)	Dissolved oxygen solubility in natural waters, calculation, multiparameter sonde.
pH, water, unfiltered, field, standard units	00400	std units	NA	NA	NA	PROBE	pH, wu, field, electrometry, multiparameter sonde (NEWSC)	Field determination of pH in unfiltered water by electrometry with multiparameter sonde.
Flow rate, instantaneous, gallons per minute	00029	gal/min	NA	NA	NA	G0009	Instantaneous flow rate, gallons per minute	Field measured flow rate; timed measurement of volume.

1 This report contains CAS Registry Numbers, which is a Registered Trademark of the American Chemical Society. CAS recommends the verification of the CASRNs through CAS Client Services.

<sup>&</sup>lt;sup>2</sup>Hach DR/2010 Spectrophotometer Procedures Manual, Hach Company (2000).

**Table 3.** Constituents, field parameters, arsenic speciation, and measuring entities for water samples collected from domestic wells in Maine, 2001–2 and 2006–7.

[USGS, U.S. Geological Survey; Maine HETL, Maine Department of Human Services Health and Environmental Testing Laboratory, Augusta, Maine; Underwriters Laboratories, Underwriters Laboratories Inc., South Bend, Indiana; NWQL, National Water Quality Laboratory, Denver, Colorado; DO, dissolved oxygen]

	Hece	L	aboratory or me	asuring e	ntity
Constituent	USGS parameter code	Maine HETL 2001–2; 2006–7	Underwriters Laboratories 2006–7	USGS- NWQL 2001–2	USGS field measurements 2001–2; 2006–7
Anal	ytical determinations				
Alkalinity	29803		X		
Arsenic, total (unfiltered)	01002	X	X	X	
Arsenic, dissolved (filtered)	01000	X			
Fecal coliform	31616		X		
Fluoride	00951		X		
Iron, total	74010/99138		X		X
Iron, ferrous (Fe II)	99128				X
Iron, ferric (Fe III)	51279				X
Lead	52298		X		
Manganese	71883/53570		X		X
Nitrate	00630/99124		X		X
Nitrite	00615		X		
pH (lab)	00403		X		
Phosphate	99126				X
Sodium	52290		X		
Sulfate	00946/99127		X		X
Sulfide	99119				X
Total hardness	00909		X		
Turbidity	82079		X		
Uranium	63032		X		
ŀ	Field parameters				
Dissolved oxygen	00300				X
Percent DO saturation	00301				X
pH (field)	00400				X
Specific conductance	00095				X
Temperature	00010				X
Arsenic spo	eciation—field prepa	ration			
Total arsenic, raw well water					X
Filtered arsenic, raw well water					X
Arsenite [As(III)], raw well water; field ion exchange					X
Arsenate [As(V)], raw well water; field ion exchange					X
Filtered arsenic, well water, post-household filtration system					X
Arsenite [As(III)], well water, post-household filtration system					X

results for many of the same constituents, and to help inform the evaluation of the water treatment system performance in both studies.

POE and POU samples were collected for laboratory chemical analysis by following standard USGS protocols (U.S. Geological Survey, variously dated) and that of the Underwriters Laboratories Inc. (UL) DrinkWell well-water testing system. Once stable readings for temperature, specific conductance, and pH were achieved (0.25–0.75 hour), POE samples were collected. Source water for POU samples (kitchen faucet) was run continuously for approximately 20 minutes prior to sample collection. The faucet aerator on the POU faucet was removed to minimize sample aeration. Faucets were disinfected with alcohol (isopropanol) wipes prior to sample collection. Bottle types and preservatives varied depending on the analysis to be performed. A complete list of constituents and minimum reporting limits is shown in table 1. All samples collected for analysis at UL were chilled to 4 degrees Celsius and shipped overnight on the day of collection. Samples were analyzed within appropriate, analytespecific holding times, according to the laboratories' standard operating procedures. Published arsenic results are labeled as either "pre-household filtration" (POE) or "post-household filtration" (POU) samples (Culbertson and others, 2020).

#### Point-of-Entry and Point-of-Use Sample Collection for Laboratory Arsenic and Arsenic Speciation (Valence) Analysis

Household well-water samples for unfiltered arsenic, filtered arsenic (0.45-µm filter), and filtered As(III), and As(V) were collected at the POE and processed onsite prior to laboratory analysis. Laboratory analyses of arsenic, for both the 2001–2 and 2006–7 sampling periods, were performed by the Health and Environmental Testing Laboratory (HETL) of the Maine Department of Human Services<sup>5</sup> in Augusta, Maine, and are published in Culbertson and others (2020). A subset of sample replicates (paired samples) were collected during the 2001–2 sampling period and analyzed by both the HETL and the USGS National Water Quality Laboratory (NWQL). Replicate results are identified as such in Culbertson and others (2020) by the "SampleType" field. In both sampling periods, the effectiveness of household water-treatment systems in removing arsenic was assessed by comparing POU samples (post-arsenic treatment system, collected at the kitchen faucet) to raw, untreated POE samples. Throughout the 2006-7 sampling period, replicate (paired) samples were collected for preand post-arsenic treatment system assessment and analyzed by both UL and HETL. During preliminary stages of the 2006–7 sampling period, post-household treatment system (POU) samples were analyzed for filtered arsenic as well as As(III); however, there was concern that any As(III) present could be oxidized by interaction with the treatment system and thus be

reported as As(V). Consequently, the sample collection protocol was modified mid-study, whereby, an additional replicate sample for As(III) determination on the raw (POE) well water was collected. Thereafter, instead of two sample replicates, As(III) determinations on raw POE well water were performed in triplicate. All samples for As(III) determinations on raw water were collected in triplicate during the 2001–2 sample period. Except where noted, the only arsenic determination on post-household filtration (POU) water was for filtered arsenic.

#### **Arsenic Sample Preparation and Preservation**

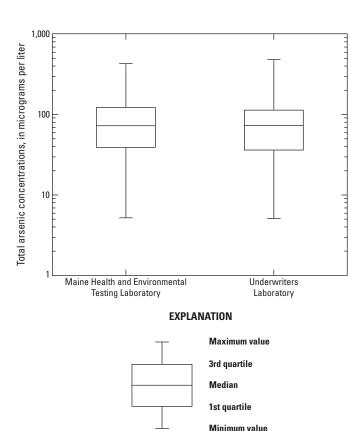
Sample preparation for arsenic analysis included acidification to pH <2 (Ultrex, ultra-high-purity nitric acid [HNO<sub>3</sub>], J.T. Baker Chemical Company, Phillipsburg, N.J.) for filtered arsenic sample analysis, and filtration through 0.45 µm nylon syringe filters, followed by acidification (5 percent [volume per volume] sulfuric acid [0.05 percent final H<sub>2</sub>SO<sub>4</sub>], Baker Instra-Analyzed, J.T. Baker Chemical Company) for filtered arsenic sample analysis and arsenic speciation (valence) analysis. Samples for As(III) and As(V) analysis were prepared onsite by eluting aliquots of the filtered/acidified samples through anion-exchange chromatography columns according to the method described by Ficklin (1983). Once aliquots for arsenic valence analysis were collected, the samples for filtered arsenic analysis were further acidified to pH <2 with HNO<sub>3</sub> (Ultrex). This step was necessary to prevent HNO<sub>3</sub> damage to the anion-exchange resin. The H<sub>2</sub>SO<sub>4</sub> step was necessary to adjust the pH of the source water to approximately pH 4. At this pH, the anion-exchange columns retain the negatively charged As(V) species while allowing the neutral As(III) species to pass through in the filtrate (Ficklin, 1983; Edwards and others, 1998; Wilkie and Hering, 1998). The filtrates were collected and further acidified to pH <2 with HNO<sub>3</sub>. Any arsenic measured in these samples is interpreted to be As(III) (table 3). As(V) was determined as the difference between filtered arsenic and As(III). Total particulate arsenic was determined as the difference between unfiltered and filtered arsenic samples.

Arsenic analyses were performed by three laboratories: (1) the Maine Department of Human Services HETL, Augusta, Maine, (2) UL, South Bend, Ind., and (3) the USGS NWQL, Denver, Colorado (table 3). Interlaboratory comparisons are displayed in figures 2 and 3 and are available in the published data (Culbertson and others, 2020).

#### Preparation of Field Anion-Exchange Chromatography Columns for Arsenic Speciation Analysis

Anion-exchange chromatography columns for the separation of As(III) and As(V) in the field were prepared in the laboratory prior to well-water sample collection. AG 1 anion-exchange resin (50–100 mesh; Bio-Rad Laboratories,

<sup>&</sup>lt;sup>5</sup>Since July 2004, the Maine Department of Health and Human Services.



**Figure 2.** Distribution of unfiltered arsenic concentrations in paired samples analyzed by the Maine Department of Human Services Health and Environmental Testing Laboratory (HETL) and Underwriters Laboratories Inc. (UL), 2006–7. Modified from Nielsen and others (2010).

Hercules, Calif.) was obtained in the chloride form and converted to the acetate form prior to column packing (Wilkie and Hering, 1998). Poly-Prep 0.8 x 4-centimeter polypropylene chromatography columns (Bio-Rad Laboratories, Hercules, Calif.) were slurry-vibra-packed with approximately 2 milliliters of the converted anion-exchange resin and excess deionized water to keep the resin moist (vibra-packing of the resin slurry was used to eliminate channeling in the resin bed). Columns were capped and refrigerated until use in the field. The anion-exchange columns were prepared in batches of 100; column efficiency tests were performed on each batch. Representative anion-exchange column efficiencies are shown in tables 4 and 5.

# Quality Assessment of Laboratory Arsenic Analysis

Arsenic in all samples was determined by inductively coupled plasma-mass spectrometry (EPA Method 200.8; U.S. Environmental Protection Agency, 1994) by the HETL (table 1). As a QA check of samples analyzed by the HETL, replicate samples, collected during 2001–2, were analyzed for unfiltered arsenic by an alternate method: USGS Method GF096: "Arsenic Determination by Hydride Generation/ Atomic Absorption Spectrometry" by the USGS NWQL (table 1).

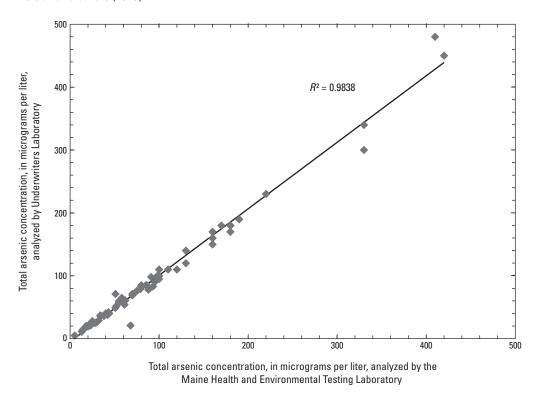


Figure 3. Arsenic concentrations in paired samples analyzed by the Maine Department of Human Services Health and Environmental Testing Laboratory and Underwriters Laboratories Inc., 2006–7. Modified from Nielsen and others (2010).

**Table 4.** Representative efficiency of anion-exchange chromatography resin for the field separation of As(III) and As(V) for single arsenic species recovery.

[As, arsenic;  $\mu$ g/L, microgram per liter As(III), arsenic-3 or arsenite; As(V), arsenic-5 or arsenate]

Replicate number	As added (μg/L)	As recoverable (μg/L)
	As(III)	
Replicate 1	62	61
Replicate 2	62	61
Replicate 3	62	57
	As(V)	
Replicate 1	67	< 0.5
Replicate 2	67	< 0.5
Replicate 3	67	< 0.5

**Table 5.** Representative efficiency of anion-exchange chromatography resin for the field separation of As(III) and As(V) for As(III)+As(V) mix recovery.

[As, arsenic; As(III), arsenic-3 or arsenite;  $\mu g/L$ , microgram per liter; As(V), arsenic-5 or arsenate]

Replicate number	As(III) added (μg/L)	As(V) added (μg/L)	As recoverable (μg/L)
Replicate 1	33	34	31
Replicate 2	33	34	32

An assessment of the HETL and UL arsenic analyses was derived from a 2006–7 comparison of arsenic determinations reported by each laboratory for replicate (paired) samples. Sixty-four samples from wells with concentrations of arsenic ranging from 5 to 420 micrograms per liter ( $\mu$ g/L) were replicated (sampled at the same time and from the same source, preserved in the same way, and analyzed by using EPA Method 200.8). The results of the analyses for unfiltered arsenic from the two laboratories were compared to assure quality in the analytical process (figs. 2 and 3; Nielsen and others, 2010; Culbertson and others, 2020).

In general, the average difference in concentration for the 64 sample pairs analyzed by HETL and UL, expressed as the absolute value of the relative percent difference between the samples, was about 7.6 percent (|[HETL-UL]/([HETL+UL]/2)|\*100). A comparison of concentrations for the paired samples (fig. 3) shows that, for the most part, this difference is primarily at the upper end of the range in concentrations of arsenic (with one exception at the lower end of the range). The range in relative percent difference is 0 to 105 percent, although only two samples have a relative percent difference greater than 16.7 percent. According to a one-way analysis of variance of the datasets, the concentration means are not significantly different at the p=0.05 level (Nielsen and others, 2010).

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