

## JOB HAZARD ANALYSIS

JOB: Core cutting

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MODIFIED: 4/25/2002 by Sarah Jablonski

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**Recommended Protective Clothing and Equipment:**

Lab coat, gloves, goggles, first aid kit, proper shoes

Sequence of Basic Job Steps	Potential Accidents/Hazards	Recommended Safe Job Procedures
Clean up lab area for cutting space (table, side table, AWMR station) and lay out white lab bench paper		
AWMR all sample vials, cutting knives, piston, core caps (for cuttings)	*See AWMR JHA *	*See AWMR JHA *
Check all vials to see they are labeled and weighed (spec. vials, cperf bags, organics)		
Fill out core processing sheet with intervals and expected subsamples	Use pencil, check for errors in interval listing	
Remove core from the reefer to be cut and drain the overlying water	Jostling sediment in the core, stuck in reefer, removing fluff layer when siphoning water	Proper PPE for being in reefer, tell someone you're going to the reefer, hold core upright and walk slowly, use care when close to sediment/water interface, pipette the last bit of water with a syringe
Find and hang the x-ray of the core being cut		
Cut the core cap with razor knife and place the piston on the bottom of the core	Cutting with razor knife, loosing sediment from bottom of core, contact with sediment	Wear proper PPE, work quickly once core cap is removed
Place core and piston on jack/cutting stand, clamp down, and jack the sediment to the top of core barrel	Core can slip in the assembly	Clamp tight, but not too tight to prevent piston movement
Borrow camera and take pictures of the top of the core and the length of the core (with a photo label)		
Tape ruler to the core and record the bottom of the piston (beginning depth)		
CUTTER 1: Jack sediment to first interval		
Cut off the edges in contact with the core barrel, scrape into AWMR core cap		
Sequence of Basic Job Steps	Potential Accidents/Hazards	Recommended Safe Job Procedures
Describe sediment interval, slice sediment into spec. vial (be sure to		

take only sediment in the interval, not below), and homogenize the sample		
Remove a cperf aliquot (about 4g wet) into weighed bag.		
CUTTER 1 Repeat steps until core is "on piston" (last interval)		
CUTTER 2: Record weight of sample+bag and bag weight to lab notebook		
Record sample+vial and vial weights in lab notebook (weighed twice)		
Describe and record color of sample, depth interval, time of day, and subsamples taken (on core processing sheet)		
Place all weighed and described samples in cooler (with blue ice) until core is finished.		
CUTTER 1: Repeat steps until core is "on piston" (last interval)		
CUTTER 3: Acid wash cutting knives, core caps, piston, and all tools in contact with sediment	*See AWMR JHA *	*See AWMR JHA *
Remove piston from finished barrel, wash barrel in soap and water and store in lab van (or at MOF)		
Dispose of core cuttings and clean up cutting area		
Store weighed specimen vials in "new" freezer (labeled crate), cperf samples in lab reefer (labeled bag), and biota in "new" reefer		

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