



# **Complexity of Nearshore Strontium-to-Calcium Ratio Variability in a Core Sample of the Massive Coral *Siderastrea siderea* Obtained in Coral Bay, St. John, U.S. Virgin Islands**

By Christopher D. Reich, Ilsa B. Kuffner, T. Don Hickey, Jennifer M. Morrison, and Jennifer A. Flannery

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## Contents

Abstract .....	1
Introduction .....	1
Setting .....	3
Methods.....	4
Coral Collection.....	4
X-Rays and Extension Rates .....	6
Sr/Ca Analysis .....	7
Results and Summary .....	8
Acknowledgments .....	10
References Cited .....	11

## Figures

1. Satellite image of St. John, U.S. Virgin Islands, shows location of coral cores collected in March 2012 .....	3
2. Photographs of a U.S. Geological Survey scientist coring a <i>Siderastrea siderea</i> colony at the U.S. Virgin Islands using a hydraulic-powered drill (A) and extracting the core (B). The Harbor Point (HP-2) coral in St. John, Coral Bay, is shown prior to coring (C) and after coring (D) .....	5
3. X-ray of the HP-2 coral slab. An example of a high-density corallite wall that would provide a good path for micro-sampling is identified by arrows.....	7
4. Plot of strontium and calcium (Sr/Ca) for Path 1 (red) and Path 2 (blue) collected from HP-2. Sr/Ca shifts colder (higher Sr/Ca) in the zone where the brown banding occurs .....	9
5. Scanned image of the HP-2 coral slab shows drilling Path 1 and a prominent brown-band layer (A) and a strontium/calcium (Sr/Ca) plot of Path 2 overlain on X-ray (B) .....	10

## Tables

1. Collection information for all corals cored in nearshore waters of St. Thomas and St. John, U.S. Virgin Islands.....	6
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## Conversion Factors

### Inch/Pound to SI

Multiply	By	To obtain
Length		
inch (in.)	2.54	centimeter (cm)
inch (in.)	25.4	millimeter (mm)
foot (ft)	0.3048	meter (m)
mile (mi)	1.609	kilometer (km)
mile, nautical (nmi)	1.852	kilometer (km)
Area		
square mile (mi <sup>2</sup> )	2.590	square kilometer (km <sup>2</sup> )

### SI to Inch/Pound

Multiply	By	To obtain
Length		
centimeter (cm)	0.3937	inch (in.)
millimeter (mm)	0.03937	inch (in.)
meter (m)	3.281	foot (ft)
kilometer (km)	0.6214	mile (mi)
kilometer (km)	0.5400	mile, nautical (nmi)
Area		
square kilometer (km <sup>2</sup> )	0.3861	square mile (mi <sup>2</sup> )

Temperature in degrees Celsius (°C) may be converted to degrees Fahrenheit (°F) as follows:

$$^{\circ}\text{F}=(1.8\times^{\circ}\text{C})+32$$

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## Abstract

Strontium-to-calcium ratios (Sr/Ca) were measured on the skeletal matrix of a core sample from a colony of the massive coral *Siderastrea siderea* collected in Coral Bay, St. John, U.S. Virgin Islands. Strontium and calcium are incorporated into the coral skeleton during the precipitation of aragonite by the coral polyps and their ratio is highly temperature dependent. The robustness of this temperature dependence makes Sr/Ca a reliable proxy for sea surface temperature (SST). Details presented from the St. John *S. siderea* core indicate that terrestrial inputs of sediment and freshwater can disrupt the chemical balance and subsequently complicate the utility of Sr/Ca in reconstructing historical SST. An approximately 44-year-long record of Sr/Ca shows that an annual SST signal is recorded but with an increasing Sr/Ca trend from 1980 to present, which is likely the result of runoff from the mountainous terrain of St. John. The overwhelming influence of the terrestrial fingerprint on local seawater chemistry makes utilizing Sr/Ca as a SST proxy in nearshore environments very difficult.

## Introduction

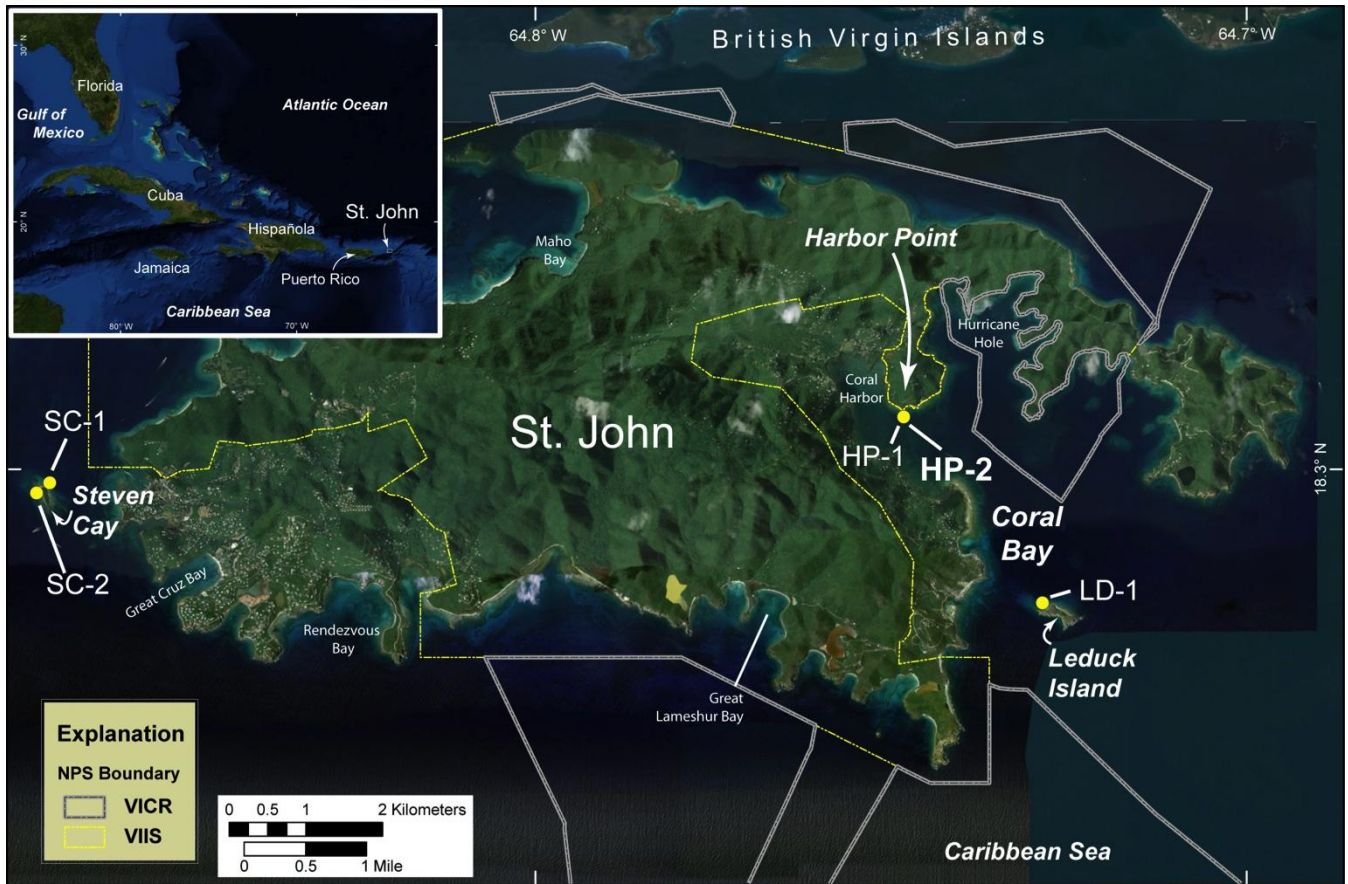
Coral reefs generally occur between 35° N and 32° S, where water temperature stays within a range from approximately 18 to 31 °C (Druffel, 1997; Veron, 2000; Corrège, 2006). This relatively narrow range in sea surface temperature (SST) tolerance allows corals to be useful as indicators of paleoclimate. Many of the shallow-water coral reefs (< 25-meter (m) water depth) in the Caribbean Sea and tropical Atlantic Ocean have been accumulating since 4,000 to 6,000 years ago when rising sea level flooded the shallow banks where modern-day corals are found (Lighty and others, 1978; Toscano and Lundberg, 1998; Balsillie and Donoghue, 2004).

Current understanding of climate variability has been limited by the short duration of instrumental temperature records. Since corals can live for long periods of time and grow in areas that are relatively consistent with respect to temperature and salinity, they are ideal for retrospectively extending instrumental SST records. Corals secrete skeletons composed of calcium carbonate (CaCO<sub>3</sub>), in the form of the mineral aragonite, through a process known as skeletogenesis (Beck and others, 1992; Druffel, 1997; Corrège, 2006; Lough and Cooper, 2011). During the secretion of aragonite, corals substitute trace elements such as barium (Ba), lead (Pb), magnesium (Mg), manganese (Mn), and

strontium (Sr) in place of calcium (Ca) (Druffel, 1997; Corrège, 2006). Studies on skeletal strontium-to-calcium (Sr/Ca) suggest that the variability of Sr in the aragonite is highly dependent upon the temperature of seawater (Weber, 1973; Smith and others, 1979; Beck and others, 1992; Alibert and McCulloch, 1997; Swart and others, 2002; Corrège, 2006). The warmer the water, the less Sr is incorporated into the aragonite matrix; therefore, lower Sr/Ca equates to warmer SST and higher Sr/Ca to cooler SST (Weber, 1973; Smith and others, 1979). Corals also deposit annual high- and low-density bands, akin to tree rings, which are useful in age-dating the coral (Lough, 2008; Lough and Cooper, 2011). The measurement from one high-density band to the next is the annual linear extension rate, which can vary from year to year due to various environmental factors, such as thermal (cold or hot) stress, disease, water quality, or predation (fish or boring organisms).

The application of Sr/Ca to paleothermometry is not without caveats. Modern-day technology and precise measurement of Sr and Ca have reduced the errors of past studies; however, there are other factors that can affect Sr and Ca concentration such as species-specific partition coefficients, metabolism, growth rate, other biological parameters known as “vital effects,” sampling techniques, and local seawater Sr/Ca variability (Smith and others, 1979; DeVilliers and others, 1994; Swart and others, 2002; DeLong and others, 2011, Kuffner and others, 2012). The goal of Sr/Ca thermometry is to measure the elemental concentration of Sr and Ca obtained from high-resolution sampling (closely spaced samples) of the coral skeleton and calibrate the Sr/Ca results with local surface water temperature data so that a calibration equation can be calculated. This assumes that local seawater Sr/Ca is conservative and stable. The calibration equation is then applied to the entire coral Sr/Ca record to calculate historical SST.

Coral cores from the U.S. Virgin Islands were collected for use in retrospective studies on climate variability using strontium-to-calcium ratios as a proxy for SST. Collection of cores from the U.S. Virgin Islands (St. Thomas and St. John) provided specimens from a tropical region of the Caribbean Sea, which will be compared to existing results from coral cores of the same species collected from the subtropical region in the Florida Keys (U.S.A.). This document reports on Sr/Ca data obtained from a core sample from a single colony of the massive coral *Siderastrea siderea* collected from Harbor Point (Coral Bay) in eastern St. John, U.S. Virgin Islands (fig. 1).



**Figure 1.** Satellite image of St. John, U.S. Virgin Islands, shows location of coral cores collected in March 2012. The *Siderastrea siderea* HP-2 (Harbor Point) core, discussed in this report, is located between Coral Harbor and Coral Bay along the eastern shore of St. John. NPS, National Park Service; VICR, Virgin Islands Coral Reef National Monument; VIIS, Virgin Islands National Park.

## Setting

St. John is located at approximately 18.3° N, 64.8° W, in the island arc chain of the eastern Caribbean Sea (fig. 1). The island of St. John encompasses an area of ~52 square kilometers and is mountainous with a maximum elevation of 380 m (Rankin, 2002; Hall and KellerLynn, 2010). The population of St. John is approximately 4,170 (2010 census), but more than 400,000 tourists visit the island annually (Hall and KellerLynn, 2010). The Virgin Islands National Park (VIIS), established in 1956, covers over 60 percent (7,000 + acres) of St. John (fig. 1; <http://www.nps.gov/viis/>). In 2001, the VIIS expanded to include a 3-mile-wide belt of submerged land, and the Virgin Islands Coral Reef National Monument (VICR) was established (Hall and KellerLynn, 2010; <http://www.nps.gov/vicr/>).

The geology of the U.S. Virgin Islands is composed of the Lameshur Volcanic-Intrusive Complex of the Cretaceous Period (Rankin, 2002). This geologic complex consists of basalts, andesite, and minor calcareous rocks and cherts that were uplifted and tilted as the Caribbean Plate moved from the western Pacific to its current position (Rankin, 2002). Though the Virgin Islands are situated within a

seismically active region, recent activity has subsided, and, unlike Cuba or Puerto Rico, there is no evidence of uplift or fault scarps since the late Pleistocene Epoch (Rankin, 2002).

The Harbor Point (HP-2) core was collected along the eastern end of St. John, within Coral Bay, off a projecting land mass known as Harbor Point (fig. 1). Coral Bay is a large, multi-lobed embayment that is fringed by mangroves, contains fringing coral reefs, and is home to ~634 people (2010 census; <http://www.nps.gov/viis/>). Both Coral Bay, south of Harbor Point, and Coral Harbor, north of Harbor Point, have been subjected to increased sedimentation over the past several decades due to road construction, residential and commercial construction, and other anthropogenic activities (Brooks and others, 2007; Gray and others, 2012). Enhanced sedimentation occurs as a result of these land-use changes during large precipitation events, such as hurricanes. Bottom sediments transition from terrigenous (volcanic derived) at the head of Coral Harbor, to open marine (carbonate derived) near the center of Coral Bay (Brooks and others, 2007). Harbor Point is in the transition zone for the mixing of terrigenous and carbonate sediments.

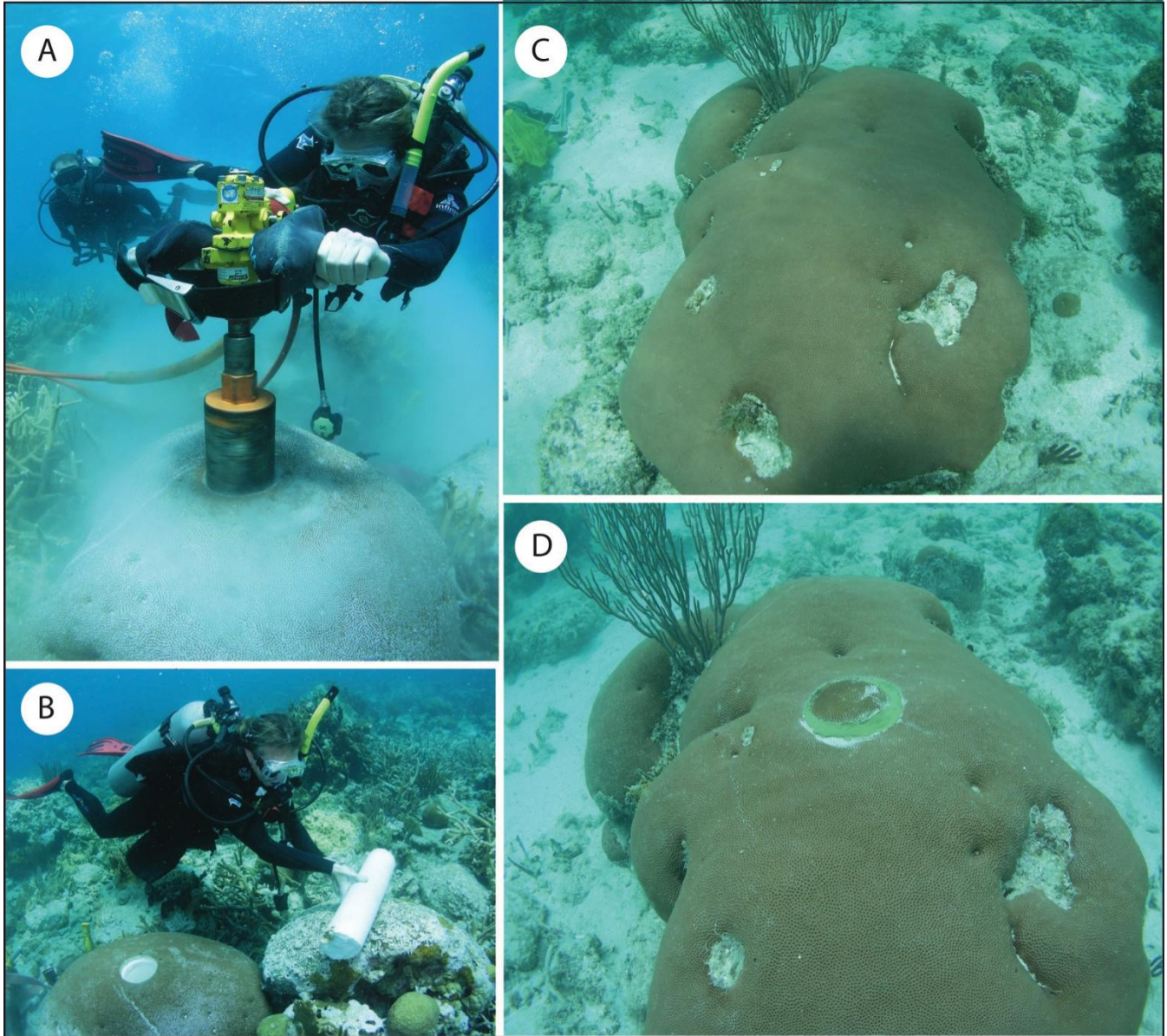
## Methods

### Coral Collection

Site selection and identification of potential coral colonies for sampling occurred in September 2011, at which time thermographs were deployed near prospective, healthy, massive corals. Thermographs were deployed in water depths <5 m at locations near Flat Cays in St. Thomas and near Leduck Island and Harbor Point in St. John.

These sites were revisited in March 2012. The largest and healthiest corals from two species, *Siderastrea siderea* and *Montastraea faveolata*, were identified at the prospective locations and cored along the primary growth axis. In addition, a single *Diploria strigosa* colony was cored near Steven Cay on the western shore of St. John (fig. 1). In total, eight coral heads were cored from the three species (see table 1 for core statistics and Reich and others, 2012, for further details). Corals were drilled using a Stanley DL07 underwater hydraulic drill and 4-inch-diameter core barrel (fig. 2A) using similar methods as Reich and others (2009) and Hickey and others (2012). A single core was collected from each coral head (fig. 2B). Cores were collected at three locations on St. John: Steven Cay, Leduck Island, and Harbor Point (fig. 1). This report, however, only discusses the HP-2 core collected from Harbor Point located in Coral Bay, St. John. The HP-2 core was collected from the massive coral *S. siderea* (fig. 2C, table 1).





**Figure 2.** Photographs of a U.S. Geological Survey scientist coring a *Siderastrea siderea* colony at the U.S. Virgin Islands using a hydraulic-powered drill (A) and extracting the core (B). The Harbor Point (HP-2) coral in St. John, Coral Bay, is shown prior to coring (C) and after coring (D). A live plug was inserted and epoxied into place after the core was extracted (D). Plug diameter is 10.2 centimeters.

**Table 1.** Collection information for all corals cored in nearshore waters of St. Thomas and St. John, U.S. Virgin Islands. Details for the Harbor Point (HP-2) core collected on St. John are discussed in this report.

[Dates are shown by month/day/year. USVI, U.S. Virgin Islands; N, north; W, west; cm, centimeter; m, meter]

<b>Date</b>	3/16/12	3/17/12	3/17/12	3/19/12	3/20/12	3/20/12	3/21/12	3/21/12
<b>Site ID</b>	FC-1	FC-2	FC-3	LD-1	HP-1	HP-2 <sup>a</sup>	SC-1	SC-2
<b>Location</b>	Flat Cays	Flat Cays	Flat Cays	Leduck Island	Harbor Point	Harbor Point	Steven Cay	Steven Cay
<b>USVI island</b>	St. Thomas	St. Thomas	St. Thomas	St. John	St. John	St. John	St. John	St. John
<b>Latitude (N)</b>	18.3166	18.3162	18.3161	18.3175	18.3399	18.3399	18.3316	18.3306
<b>Longitude (W)</b>	64.9882	64.9883	64.9882	64.6902	64.7054	64.7054	64.807	64.8082
<b>Total core length (cm)</b>	77.5	60.3	100.3	35.6	165.1	92.1	80.0	54.6
<b>Water depth (m)</b>	4.85	4.24	4.54	3.63	2.72	3.93	3.63	0.3
<b>Coral species</b>	<i>S. siderea</i>	<i>S. siderea</i>	<i>M. faveolata</i>	<i>S. siderea</i>	<i>M. faveolata</i>	<i>S. siderea</i>	<i>S. siderea</i>	<i>D. strigosa</i>

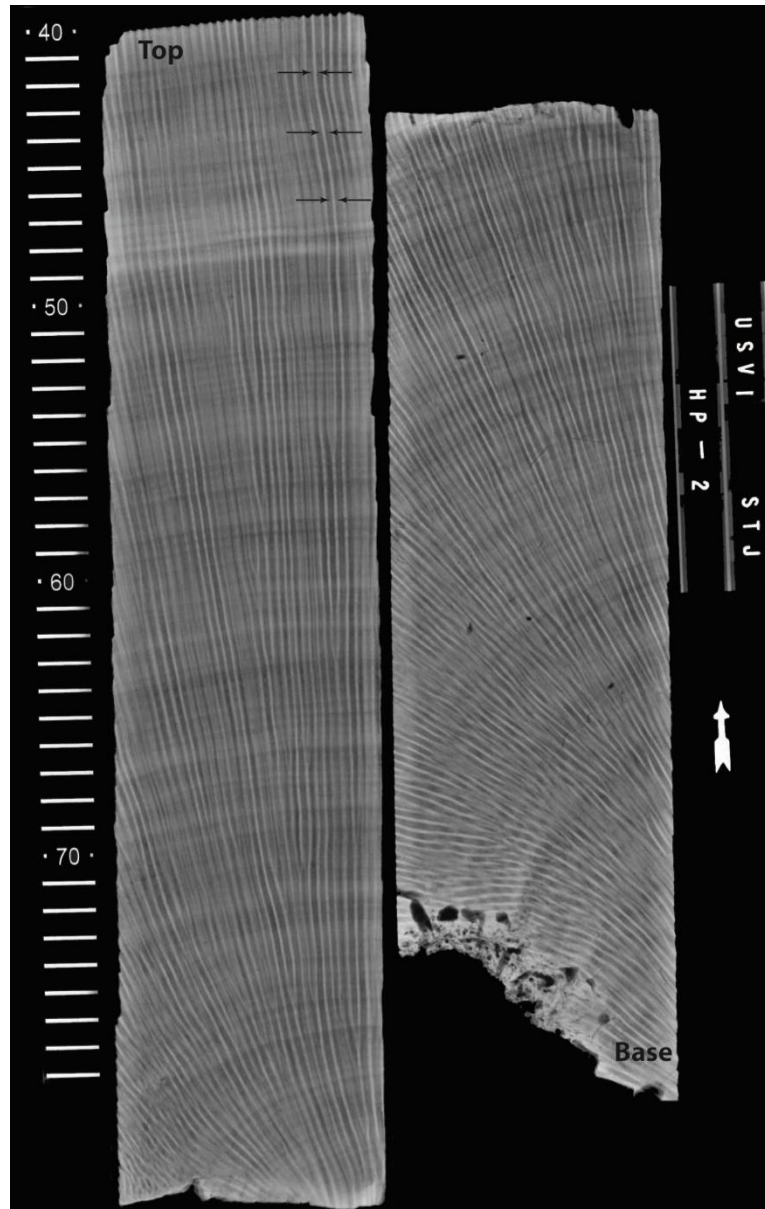
<sup>a</sup>Presented in this report.

A short plug with live coral tissue was collected from the base of the coral head and inserted into the open hole where the core was extracted. This method, similar to that described by Hudson (1983), was implemented in order to speed up the recovery time of the coral. The short plug from the base was placed in the hole and epoxied into place (fig. 2D). In most situations the dense skeletons of *S. siderea* and *D. strigosa* complicated and prevented the live tissue core plug transplant method. Since the live tissue plug could not be used for many of the *S. siderea* and *D. strigosa* corals, a precast cement plug was epoxied within the hole so that no live tissue edges were left exposed.

## X-Rays and Extension Rates

Cores were sectioned along the primary growth axis into 4-millimeter-thick slabs. Multiple (~3) slabs were obtained because corallite walls tend to migrate over time and therefore one slab may have a more continuous path for geochemical sampling. Slabs were placed in deionized water and ultrasonicated using a Branson Sonifier with probe to remove fine-grained material. Slabs were allowed to completely dry prior to being X-rayed. The HP-2 slab identified for sampling was placed on a phosphor plate and X-rayed at 55 kilovolts and 2.5 milli-Ampere-seconds (mAs). The distance between the plate and the X-ray source was 79 centimeters (cm). The plate was scanned on an iCR3600+ scanner at 254 dots per inch (dpi) resolution (10 pixels mm<sup>-1</sup>) and processed on iCRco, Inc. software. The X-ray (fig. 3) was exported as both 16-bit Tagged Image File Format (TIFF) and native Digital Imaging and Communications in Medicine (DICOM) file formats.

X-ray images were examined for high- and low-density bands. A couplet of high- and low-density bands (light and dark) on the X-ray is equivalent to one year of growth. Adobe Photoshop was used to measure the distance between couplets. An estimate of annual growth, or linear extension, rate over the length of the coral slab was calculated (Hudson and others, 1976; Helmle and others, 2011). Extension rate calculations are necessary to calculate a proper sampling interval for coral powder collection via a micro-drill. A minimum of 12 samples per year are necessary to potentially capture the annual seasonal cycle in Sr/Ca. However, there is a trade-off between number of samples per year and quantity of material; increased sampling rate per year equates to less coral material per sample for Sr/Ca analyses.



**Figure 3.** X-ray of the HP-2 coral slab. An example of a high-density corallite wall that would provide a good path for micro-sampling is identified by arrows. Note faint high- and low-density bands on the X-ray. Scale is in centimeters.

### Sr/Ca Analysis

The X-ray image of the HP-2 coral slab was used to determine the appropriate path for micro-sampling. Generally, the best path to sample is the longest continuously growing corallite wall that appears as a longitudinal high-density band on the X-ray. The drilling path was traced along the corallite wall using Adobe Illustrator (fig. 3). The traced path was exported from Adobe Illustrator as a Drawing Exchange Format (DXF) file and imported into the SuperCam software operating the XYZ drill.

A 1 millimeter (mm) tungsten carbide-tipped drill bit was used to sample the coral. Coral powder was collected along the growth axis in increments of 0.25 mm (Path 1) and 0.20 mm (Path 2) to a depth of 1.25 mm, then placed in pre-labeled micro-centrifuge tubes. Based on X-ray-determined linear extension rates, approximately 12 to 16 samples per year were collected at this sampling increment. A divot in the skeleton was made every 20 samples for future referencing and is visible on subsequent X-ray images.

Sr and Ca were measured on a PerkinElmer 7300 Dual View Inductively Coupled Plasma Optical Emission Spectrometer (ICP-OES) at the U.S. Geological Survey St. Petersburg Coastal and Marine Science Center (SPCMSC). For sample analyses, ~100 micrograms ( $\mu\text{g}$ ) of coral powder was weighed, placed into an ICP tube, and acidified with 2 percent nitric acid ( $\text{HNO}_3$ ) to an approximate concentration of 20 parts per million (ppm) Ca. To measure precision and correct for potential instrumental drift, an internal gravimetric standard (IGS) solution was measured for Sr/Ca before and after each dissolved coral sample (Schrag, 1999). The average corrected IGS precision for Sr/Ca was 0.009 millimoles per mole (mmol/mol) ( $1\sigma$ ,  $n=205$ ). A second standard—homogenized powder from the coral *Porites lutea* (PL)—was analyzed for Sr/Ca to test for any potential matrix effects. The PL standard had an average corrected precision of 0.019 mmol/mol ( $1\sigma$ ,  $n=297$ ).

## Results and Summary

Extension rate (ER) measurements on HP-2 were difficult because the *S. siderea* coral collected from St. John deposited a very dense skeletal matrix (fig. 3). The dense composition made discerning annual banding (contrast between high- and low-density band couplets) challenging, even after making brightness and contrast adjustments to the X-ray image. Average linear extension rate, based on X-ray measurements, for the full length of the core (92.1 cm) was 0.40 centimeters per year (cm/yr) ( $n=155$ ). This ER dates the coral base to approximately the year 1782 (~230 years old).

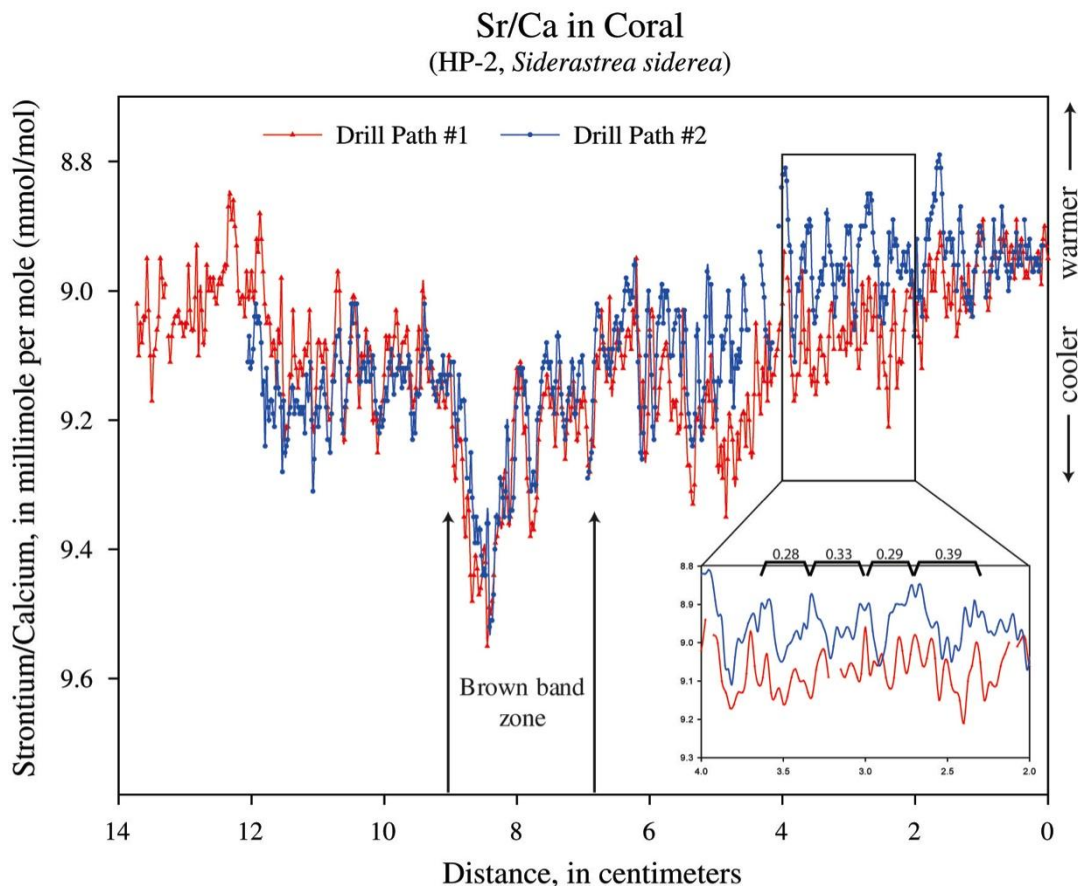
In addition to measuring high- and low-density band couplets on X-rays, we measured the distance between the presumed annual peaks on the Sr/Ca curve. Extension rates based on X-ray measurements for the top 12 cm ranged from 0.32 to 0.79 cm/yr with an average of 0.51 cm/yr ( $n=24$ ). At 12 cm from the top of the core, the last sample would have been collected at ~1989 (~23 years ago). In contrast, ER based on Sr/Ca peaks, for the top 12 cm, ranged from 0.12 to 0.46 cm/yr, with an average of 0.26 cm/yr ( $n=45$ ). The sampling period, based on ER calculated from Sr/Ca peaks, would therefore range from 2012 to ~1968 (~44 years).

The difference in average ER (0.21 cm/yr) between the two methods is cause for concern because sampling rate is based on these values, where minimum sampling density is 12 samples per year and ideal is ~20 samples per year. The slower linear extension rates, based on the Sr/Ca measurements, indicate that the goal of obtaining a minimum of 12 samples per year was marginally achieved and that by recalculating the actual average sampling rate, we only managed to sample at ~10 samples per year for Path 1 (0.25 mm/sample) and ~13 samples per year (0.20 mm/sample) for Path 2. Both of these sampling rates are at the minimal range for capturing full seasonal range in the Sr/Ca record.

Two overlapping paths were drilled to confirm Sr/Ca results and to observe any variation in results from changing the sampling rate. Annual Sr/Ca cyclicity is best captured in the Sr/Ca record from Path 2, but its continuity over time is influenced (controlled) by external anthropogenic sources (figs. 4, 5). A typical open-ocean Sr/Ca record from *S. siderea* in the Dry Tortugas (Florida Keys, U.S.A.) has no mean shift and annual cycles are distinct and highly correlative to SST (for example, see DeLong and others, 2011). Path 2, with a higher sampling resolution than Path 1, shows a slightly more prominent



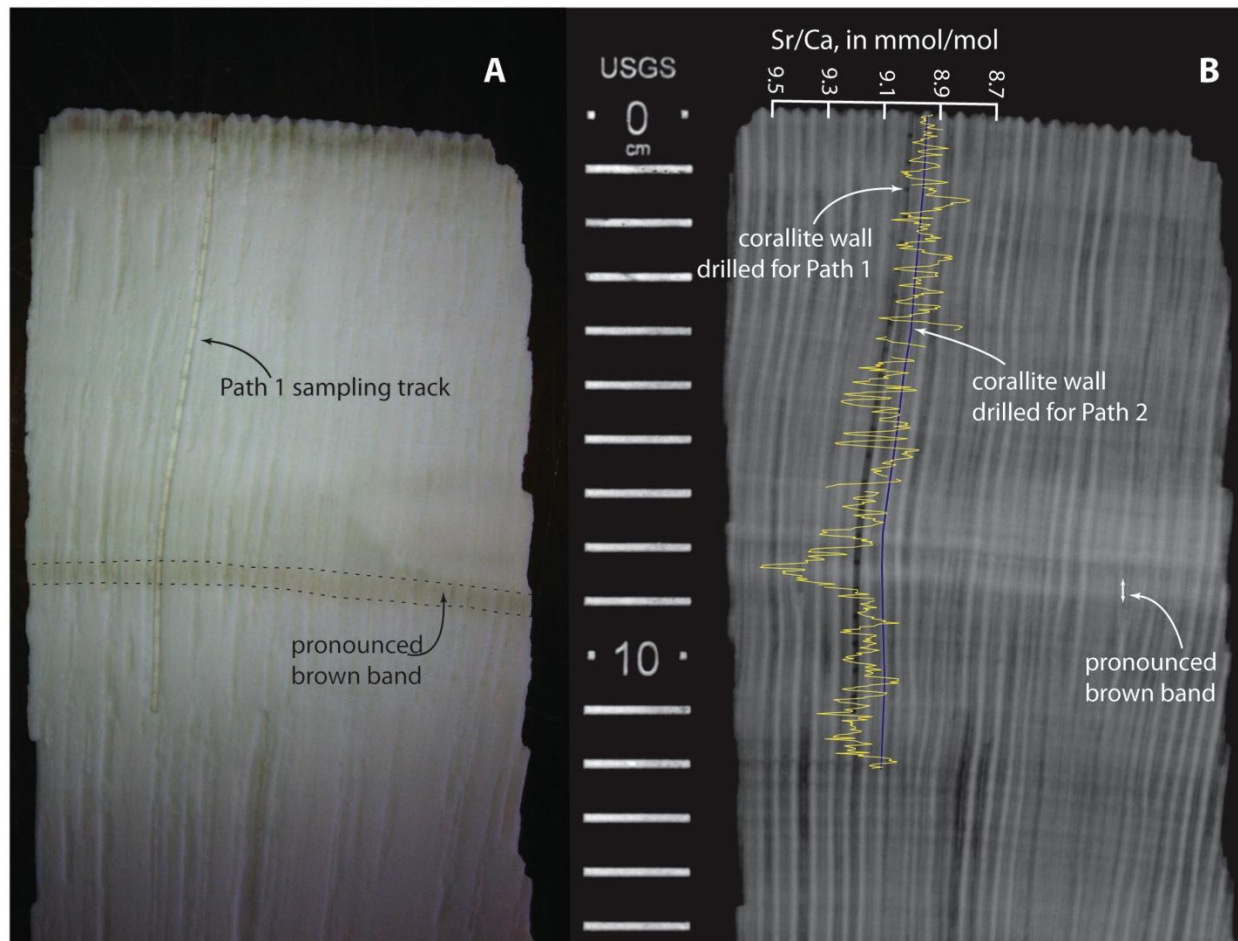
annual cycle in Sr/Ca. Annual Sr/Ca cycles for both sampling paths indicate a cool water signal (higher Sr/Ca) that began in ~1980 at the start of the brown band and thereafter begin to show SST warming to the present (figs. 4 and 5B).



**Figure 4.** Plot of strontium and calcium (Sr/Ca) for Path 1 (red) and Path 2 (blue) collected from HP-2. Sr/Ca shifts colder (higher Sr/Ca) in the zone where the brown banding occurs. Bottom right graph is an expanded view of the 2- to 4-centimeter interval that shows the annual cycles of Sr/Ca. The distance between annual cycles (values in centimeters per year shown above the brackets) for Path 2 (blue) represents the linear extension rate for HP-2.

Sr/Ca results from HP-2 show the complexity of using Sr/Ca as an SST proxy in the nearshore environment (fig. 5). Utilization of Sr/Ca as a proxy for SST is ideal for locations possessing few extraneous inputs such as terrestrial discharge of sediments and freshwater (DeLong and others, 2011). HP-2 is located in Coral Bay, near the mouth of one of the largest catchment areas on St. John. During heavy rainfall events, runoff of sediment-laden water flows into Coral Harbor and into the northern portions of Coral Bay (Brooks and others, 2007). These inputs can influence the chemistry of the surface water, impacting the Sr/Ca values of the deposited aragonite skeletal matrix. In addition, sediment is also occasionally incorporated in the skeleton, as seen in HP-2 (fig. 5A). The brown band that occurs between 7 and 9 cm down core was most likely the result of a severe runoff event leading to very turbid waters inundating the corals for several days. Assuming this scenario, the sediment would have been heavy enough to blanket the corals for an extended period of time but not severe enough to result in complete

mortality. Over time the sediment would then have been incorporated into the skeletal matrix. Results from the HP-2 core indicate that Sr/Ca is a challenging proxy for retrospective SST studies in nearshore corals because of the complexities in surface-water chemistry due to the influence of terrestrial runoff.



**Figure 5.** Scanned image of the HP-2 coral slab shows drilling Path 1 and a prominent brown-band layer (A) and a strontium/calcium (Sr/Ca) plot of Path 2 overlain on X-ray (B). Plotting the Sr/Ca on the X-ray shows the large deviation within the brown-band layer. Scale is in centimeters; millimoles per mole is shown as mmol/mol.

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