



Soil Data for a Vegetation Gradient Located at Bonanza Creek Long Term Ecological Research Site, Interior Alaska

By Kristen L. Manies, Jennifer W. Harden, Christopher C. Fuller, Xiaomei Xu, and John P. McGeehin

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Conversion Factors

International System of Units to U.S. customary units

Multiply	By	To obtain
Length		
centimeter (cm)	0.3937	inch (in.)
millimeter (mm)	0.03937	inch (in.)
meter (m)	3.281	foot (ft)
meter (m)	1.094	yard (yd)
Area		
square meter (m ²)	10.76	square foot (ft ²)
Mass		
gram (g)	0.03527	ounce, avoirdupois (oz)
kilogram (kg)	2.205	pound avoirdupois (lb)

Temperature in degrees Celsius (°C) may be converted to degrees Fahrenheit (°F) as $^{\circ}\text{F} = (1.8 \times ^{\circ}\text{C}) + 32$.

Datum

Horizontal coordinate information is referenced to the World Geodetic System of 1984 (WGS84).

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Abstract

Boreal soils play an important role in the global carbon cycle owing to the large amount of carbon stored within this northern region. To understand how carbon and nitrogen storage varied among different ecosystems, a vegetation gradient was established in the Bonanza Creek Long Term Ecological Research (LTER) site, located in interior Alaska. The ecosystems represented are a black spruce (*Picea mariana*)–feather moss (for example, *Hylocomium* sp.) forest ecosystem, a shrub-dominated ecosystem, a tussock-grass-dominated ecosystem, a sedge-dominated ecosystem, and a rich fen ecosystem. Here, we report the physical, chemical, and descriptive properties for the soil cores collected at these sites. These data have been used to calculate carbon and nitrogen accumulation rates on a long-term (decadal and century) basis (Manies and others, in press).

Introduction

High-latitude soils contain approximately 50 percent of the carbon (C) within the global soil C pool (Kasischke, 2000; Tarnocai and others, 2009) and, therefore, play an important role in the global C cycle. Climate change is expected to impact the boreal region in many ways (Hinzman and others, 2005), which in turn will affect the balance of C inputs and losses to the soil C pool. It is also important to understand nitrogen (N) stocks and availability because N controls many aspects of plant productivity and, therefore, cycling of C and N are closely linked (Vile and others, 2014).

To help our understanding of C and N stocks in a variety of boreal ecosystems, we measured physical, chemical, and descriptive soil properties in five different ecosystems in interior Alaska. These ecosystems were situated along gradients of soil moisture and dominant vegetation. Factors such as parent material, climate, and topography, which influence soil formation (Jenny, 1941), were consistent along these gradients. We used these data to calculate C and N accumulation rates on both decadal and century timescales to determine how the interaction of soil and vegetation influences these rates, and thus, C and N storage over time (Manies and others, in press). Findings from this study indicate that the C and N dynamics of the

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rich fen are different from the other four ecosystems, likely the result of differences in nutrient cycling and the fire return interval.

Site Description

Study sites were located on State land adjacent to the U.S. Forest Service Bonanza Creek Experimental Forest, approximately 30 km southwest of Fairbanks, Alaska, near the floodplain of the Tanana River (lat 64.696° N., lon 148.321° W.). These sites were part of the network of sites in the Bonanza Creek Long Term Ecological Research (LTER) program. We studied soils in five ecosystems located along a ~300-m transect. Each ecosystem was dominated by a different type of vegetation: (1) a closed-canopy black spruce (*Picea mariana*) forest with a feather moss (for example, *Hylocomium* sp.) and ericaceous shrub understory (herein, “black spruce”); (2) a shrub system consisting of willow (*Salix* sp.) and birch (*Betula* sp.) with an understory dominated by leatherleaf (*Chamaedaphne calyculata*) and sparse moss cover (herein, “shrub”); (3) a tussock-grass system dominated by *Calamagrostis canadensis* with some brown mosses present (herein, “tussock grass”); (4) a peatland dominated by emergent vegetation such as sedges (herein, “sedge”); and (5) a moss-dominated rich fen, consisting of both *Sphagnum* sp. and brown mosses (herein, “rich fen”). Several publications describe other data obtained from these sites, including ecosystem respiration (McConnell and others, 2013) and microbial abundance (Waldrop and others, 2012).

Soil Sampling

Three soil cores, encompassing all of the organic soil and extending into the mineral soil below, were collected at each site at randomly selected locations within an area of less than ~10 m². Sampling for the black spruce and low shrub site occurred during the summer and samples were obtained using a combination of soil blocks cut to a known volume and a Makita coring device (4.8 cm diameter; Nalder and Wein, 1998). Soil cores from the other three sites were obtained in the spring, when the ground was frozen, using a SIPRE corer (7.6 cm diameter; Rand and Mellor, 1985).

Each soil profile was then divided into subsamples representing soil horizons and (or) subsections of different soil horizons. This separation occurred either in the field or, if frozen, in the lab, based on visual factors such as level of decomposition and root abundance. These horizon samples were described using modified soil survey techniques following the U.S. Department of Agriculture’s Natural Resource Conservation Service (USDA-NRCS; Soil Survey Staff, 1998) and Canadian (Soil Classification Working Group, 1998) methodologies. We modified horizon codes according to the following scheme:

- L Live moss, which is green and generally contains some leaf and needle litter.
- D Dead moss, which is undecomposed or slightly decomposed. This horizon is the upper fibric organic horizon that contains more moss than roots. This horizon would be considered an O_i horizon (U.S. soil system).
- F Fibric (Canadian soil system) or fibrous organic horizon, which varies in the degree of decomposition, but in which roots are more abundant than recognizable plant parts. This horizon would be considered an O_i horizon (U.S. soil system).
- M Mesic (Canadian soil system) organic horizon, which is moderately decomposed, with few, if any, recognizable plant parts other than roots. This horizon is generally considered an O_e horizon (U.S. soil system).

H Humic (Canadian soil system) or sapric organic horizon, which is highly decomposed. The soil in this horizon smears when rubbed and contains no recognizable plant parts. The H horizon is generally considered an O_a horizon (U.S. soil system).

A Upper mineral soil that forms at the surface or just below organic soil horizons (U.S. and Canadian soil systems), which contains less than 20-volume-percent organic matter, as judged in the field.

B Lower mineral soil that has formed below an A horizon (U.S. and Canadian soil systems), with little or none of its original lithologic structure.

GR Horizon of matted grass (mostly dead, but some live) at the soil surface.

LT Litter horizon (dead leaves, twigs).

LW Liverwort.

X Unknown (not described).

Each soil horizon was labeled with a four-letter code representing the site, core, and basal depth of the sample. The first two letters of the site code, BZ, denoted that the samples were from the Bonanza Creek LTER. The next third and fourth letters represent the site from which the samples were obtained: BS, black spruce; WB, shrub site; GR, tussock-grass site; EC, sedge site; and MR, rich fen. The BZMR cores were obtained outside a plot that was part of a raised water table experiment (MR signifies manipulated-raised, see Chivers and others, 2009). After the site code, each sample was labeled with a number representing the core/soil-profile, followed by a decimal point and the basal depth of the sample (in centimeters). For example, BZWB 1.8.5 denotes a sample from the shrub site, soil profile 1, with a basal depth of 8.5 cm.

Soil-Sample Preparation and Drying

All samples were placed on open shelves in an isolated room and allowed to air-dry to a constant weight, as determined by weights measured at least 1 week apart. Temperature during air-drying ranged from 20 °C to 30 °C. After air-drying, samples were oven-dried for 48 hours in a forced-draft oven. Samples that were classified as organic soil horizons (for example, moss, litter) were oven-dried at 65 °C to avoid loss or alteration of organic matter by oxidation or decomposition. The remaining samples were oven-dried at 105 °C.

After oven-drying, samples were thoroughly mixed and split into subsamples for analysis and archiving. (Archive fractions of most of the samples described here are available by contacting K. Manies at the U.S. Geological Survey in Menlo Park, Calif.). Samples were then processed in one of two ways, depending on the horizon code. Mineral soil samples were gently crushed using a mortar and pestle, with care to break only aggregates, and then sieved through a 2-mm screen. Soil particles that did not pass through the screen were removed, weighed, and saved separately; soil that passed through the screen was then ground by using a mortar and pestle to pass through a 60-mesh (0.246-mm) screen. The ground material was mixed and placed in a labeled glass sample bottle for subsequent analyses. Organic samples were weighed, and roots wider than 1 cm in diameter were removed, weighed, and saved separately. The remaining sample material was then milled in an Udy Corp. Cyclone Sample Mill to pass through a 0.5-mm screen. The milled samples were thoroughly mixed, and a representative sample was placed in a labeled glass sample bottle for analytical chemistry.

Soil Laboratory Methods

Total Carbon, Total Nitrogen, $\delta^{13}\text{C}$ Measurements

We analyzed soil samples for total carbon and nitrogen using a Carlo Erba NA1500 elemental analyzer. Samples were combusted in the presence of excess oxygen. The resulting sample gases were carried by a continuous flow of helium through an oxidation furnace, followed by a reduction furnace, to yield CO_2 , N_2 , and water vapor. Water was removed by a chemical trap and CO_2 and N_2 were chromatographically separated before the quantification of C and N (Pella, 1990a,b). For samples that also have $\delta^{13}\text{C}$, the elemental analyzer was coupled to a Micromass Optima isotope ratio mass spectrometer (IRMS) in continuous flow mode. For these samples C and N were determined by integrating the major ion peaks (mass of 44 for CO_2 , and mass of 28 for N_2). For reliable quantification of $\delta^{15}\text{N}$, 15 to 30 μg N are generally needed and few samples met this criterion. Thus the $\delta^{15}\text{N}$ data are not reported here; these data are available from the authors by request. The $\delta^{13}\text{C}$ data are reported as deviations in parts per thousand (‰) relative to a standard, here Vienna Pee Dee Belemnite (V-PDB). $\delta^{13}\text{C}$ was calculated using the following equation:

$$\delta^{13}\text{C} \text{ ‰} = \left(\left(\frac{{}^{13}\text{C sample} / {}^{12}\text{C sample}}{{}^{13}\text{C standard} / {}^{12}\text{C standard}} \right) - 1 \right) \times 1000 \quad (1)$$

where all C measured was considered organic C because carbonates tend not to exist in the study area.

All samples were compared to a main working standard, ethylene diamine tetra-acetic acid (EDTA), for which the $\delta^{13}\text{C}$ value has been determined through a set of international standards. The chemical formula of this compound corresponds to a C concentration of 41.09 percent and N concentration of 9.59 percent. Additional working standards were analyzed as samples in all runs to check consistency and overall precision. Peach tree leaves (SRM-1547), issued by the National Institute of Standards and Technology (NIST), were included in runs composed of organic soil. A marine sediment (MESS-1), issued by the Chemistry Division of the Canadian National Research Council, was included in runs composed of mineral soils. Certified values for these standards were obtained from Becker (1990) and Govindaraju (1989), respectively. Average measured values and standard deviations of these working standards are shown in table 1.

Table 1. Statistics of analyses for working standards run on the Carlo Erba 1500, from 2006–2010. [Certified values: MESS-1, 2.99 percent carbon; SRM-1547, 2.94 percent nitrogen. Stdev., standard deviation; N, number of samples]

Standard	Percent carbon			Percent nitrogen		
	Average	Stdev.	N	Average	Stdev.	N
MESS-1	2.92	0.10	148	0.20	0.03	148
SRM-1547	46.79	0.46	177	2.84	0.13	177

Radiocarbon Measurements

A subset of bulk organic matter and picked macrofossil samples were analyzed for radiocarbon (^{14}C) to evaluate the ages of C in the soil profile. A little more than half of the samples were processed at the U.S. Geological Survey (USGS) Radiocarbon Laboratory in Reston, Va. and analyzed by accelerator mass spectrometry at the Lawrence Livermore National Laboratory Center for Accelerator Mass Spectrometry (CAMS). The other samples were sent to

the W.M. Keck Carbon Cycle Accelerator Mass Spectrometry Laboratory at the University of California, Irvine. For all samples, subsamples were combusted at 900 °C in evacuated, sealed quartz tubes in the presence of cupric oxide (CuO) and silver (Ag) wire. Following cryogenic purification, CO₂ was reduced to graphite in a reaction at 500–550 °C using the sealed-tube Zn-reduction method at the Keck laboratory (Xu and others, 2007) and the H₂-reduction method at USGS/CAMS (Vogel and others, 1984). Radiocarbon data are reported as Δ¹⁴C, fraction modern (FM), and radiocarbon age following the conventions of (Stuiver and Polach, 1977). At the Keck laboratory, the reported Δ¹⁴C values have been corrected for mass-dependent fractionation to a common value of –25 parts per thousand using the simultaneous accelerator mass spectrometry (AMS) δ¹³C measurement. At USGS/CAMS an aliquot of CO₂ from the sample combustion was independently analyzed for δ¹³C to correct for fractionation in each sample.

²¹⁰Pb, ²²⁶Ra, and ¹³⁷Cs Measurements

Dried and ground subsections of cores were analyzed for ²¹⁰Pb, ²²⁶Ra, and ¹³⁷Cs activities as measured by gamma spectrometry. We used a Princeton Gamma HPGe germanium well detector to estimate dry matter accumulation rates for assigning dates to core profiles following methods described in Van Metre and Fuller (2009) and references therein. Briefly, detector efficiency for each isotope was determined using NIST traceable standards (¹³⁷Cs = NBS SRM 4350b; ²¹⁰Pb = NBS SRM 4337; ²²⁶Ra = NBS SRM 4969). Total ²¹⁰Pb activity is the combination of supported ²¹⁰Pb (produced within the soil through the decay of ²²⁶Ra) and unsupported ²¹⁰Pb (produced in the atmosphere by radon decay and added to the ecosystem through atmospheric deposition). Unsupported ²¹⁰Pb was defined as the difference between measured total ²¹⁰Pb and ²²⁶Ra, its long-lived progenitor determined from the activities of ²¹⁴Pb and ²¹⁴Bi gamma-emitting daughters of ²²⁶Ra. Horizon subsamples from each soil profile were measured until unsupported ²¹⁰Pb was no longer detected. Unsupported ²¹⁰Pb values were then used to calculate dry mass accumulation rates (MAR, in grams per cubic centimeter per year) for each soil horizon, from which dates of formation were calculated using the Constant Rate of Supply (CRS) model (Appleby and Oldfield, 1978; these dates can be found in the Supplemental section of Manies et al., 2016). Note that for the three grass-soil profiles we did not reach the point at which unsupported ²¹⁰Pb could not be detected. Because we found ²¹⁰Pb in the mineral soil for the grass soil profiles we do not suggest using these data for dating purposes at this site. ¹³⁷Cs, an anthropogenic radionuclide (half-life of 30.1 years), provides a means to constrain sediment accumulation rates and chronology because of its well-known input history from atmospheric fallout from aboveground nuclear weapons testing in the 1950s and first half of the 1960s, with maximum deposition occurring during the years 1963 and 1964 (Callender and Robbins, 1993). Measured ¹³⁷Cs activities were decay corrected for the period of time between sample collection and analysis. Uncertainty in the measured activity of ¹³⁷Cs, ²¹⁰Pb, and ²²⁶Ra was calculated from the counting error of samples and background spectra; it's reported at the one-standard-deviation level.

Dataset Descriptions

There are three separate downloadable files containing the soil data collected from the sites described in this report. Appendix 1 lists in detail the data within the following three spreadsheet files: (1) BZ_Gradient_Field, contains field descriptions of the sampled soils, such

as root abundance, color, and soil texture; (2) BZ_Gradient_Physical contains physical descriptions of the samples, such as volumetric field moisture and bulk density; (3) BZ_Gradient_Chemistry contains elemental C, elemental N, ^{14}C , and the radioactive isotope values.

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Appendix 1. Data File Descriptions

The data files are available online only as Excel (.xlsx) spreadsheets. The zip file (.zip) containing these files is available at <http://dx.doi.org/10.3133/ofr20161034>. The content of these files is described in this appendix.

Bonanza_Gradient_Field

This file contains the field descriptions of the sampled soils. Units for each column are given on row 2 of the file. This file includes the following columns:

Sample ID	Sample identification: The first two letters in the sample ID represent the area of study, Bonanza Creek LTER (BZ). The next two letters represent the ecosystem (BS, black spruce; WB, shrub; GR, tussock grass; EC, sedge; MR, rich fen). Then there is a space. The number that follows indicates the sample number. A decimal point separates the sample number from the basal depth of the sample (in centimeters).
Depth	Indicates the basal depth, in centimeters, of sampling increment.
Field horizon code	Horizon type of the sample, as defined in the field. L, live moss; D, dead moss; F, fibric organic matter (OM); M, mesic OM; H, humic OM; A, A mineral soil horizon; B, B mineral soil horizon; GR, grass; LT, litter; LW, liverwort; X, unknown horizon. For more information about these horizon types see the Soil Sampling section of this report.
Sample description	Brief description of the sample.
Roots	Root abundance and size using conventions of USDA-NRCS (Soil Survey Staff, 1998).
Lab pH	pH value of mineral soil measured in the laboratory. Deionized water was added to create a soil slurry (50 percent soil, 50 percent water) prior to measurement.
Munsell moist color	Munsell color value according to the Munsell Soil Color Chart, measured on moist soil
Von Post or texture	If organic soil, the classification using the von Post scale of humification (Damman and French, 1987). If mineral soil, the soil texture class as described in the field, following conventions of USDA-NRCS (Soil Survey Staff, 1998).
Height above mineral	Height of each basal depth above the mineral soil boundary. Therefore, the bottom organic layer equals zero and mineral horizons are negative numbers.

Bonanza_Gradient_Physical

This file contains physical data such as bulk density and volumetric moisture content. Units for each column are given on row two of the file. Column definitions are as follows:

Sample ID	Sample identification: The first two letters in the sample ID represent the area of study, Bonanza Creek LTER (BZ). The next two letters represent the ecosystem (BS, black spruce; WB, shrub; GR, tussock grass; EC, sedge; MR, rich fen). Then there is a space. The number that follows indicates the sample number. A decimal point separates the sample number from the basal depth of the sample (in centimeters).
Depth	Indicates the basal depth, in centimeters, of sampling increment.
Field horizon code	Horizon type of the sample, as defined in the field. L, live moss; D, dead moss; F, fibric organic matter (OM); M, mesic OM; H, humic OM; A, A mineral soil horizon; B, B mineral soil horizon; GR, grass; LT, litter; LW, liverwort; X, unknown horizon. For more information about these horizon types see the Soil Sampling section of this report.
Sample description	Brief description of the sample.
Date sampled	Date the sample was taken (month/day/year).
Thickness	Thickness of soil horizon.
>2 mm in sample	Dry weight percent of soil particles not passing through a 2 mm sieve after gentle crushing.
>1 cm in sample	Dry weight percent of roots larger than 1 cm in diameter in the sample.
Bulk density (<2 mm)	Grams of oven-dried soil per cubic centimeter, with soil particles greater than 2 mm and roots greater than 1 cm diameter removed.
Bulk density	Grams of oven-dried soil per cubic centimeter for the entire soil sample with no fractions excluded. Calculated similarly to "Bulk density (<2 mm)" except the weight of particles greater than 2 mm and roots greater than 1 cm diameter have been included.
Volumetric field moisture (OD)	The percentage of water in the sample, by volume, of the oven dried (OD) sample.
Convert from AD to OD bulk density	Moisture remaining in a sample after air-drying (AD) to constant weight as determined by subsequently oven-drying (OD) the sample. Bulk density, on an air-dried basis, equals $\text{Bulk density}_{\text{oven dry}} / (1 - (\text{Convert from AD to OD bulk density}/100))$.

Bonanza_Gradient_Chemistry

This file contains analytical data for the samples. Units for each column are given on row 2 of the file. Column definitions are as follows:

Sample ID	Sample identification: The first two letters in the sample ID represent the area of study, Bonanza Creek LTER (BZ). The next two letters represent the ecosystem (BS, black spruce; WB, shrub; GR, tussock grass; EC, sedge; MR, rich fen). Then there is a space. The number that follows indicates the sample number. A decimal point separates the sample number from the basal depth of the sample (in centimeters).
Depth	Indicates the basal depth, in centimeters, of sampling increment.
Field horizon code	Horizon type of the sample, as defined in the field. L, live moss; D, dead moss; F, fibric organic matter (OM); M, mesic OM; H, humic OM; A, A mineral soil horizon; B, B mineral soil horizon; GR, grass; LT, litter; LW, liverwort; X, unknown horizon. For more information about these horizon types see the Soil Sampling section of this report.
Sample description	Brief description of the sample.
Carbon	Percentage by weight of total carbon (C) in an oven-dried soil sample with material greater than 2 mm or 1 cm diameter removed.
Nitrogen	Percentage by weight of total nitrogen (N) in an oven-dried soil sample with material greater than 2 mm or 1 cm diameter removed.
$\delta^{13}\text{C}$	Stable isotope signature of oven-dried sample with material >2 mm or 1 cm diameter removed.
LOI	Loss-on-ignition value of oven-dried sample with material >2 mm or 1 cm diameter removed.
^{137}Cs	Amount of ^{137}Cs measured in the sample.
^{137}Cs error	Laboratory error associated with the ^{137}Cs measurement.
^{210}Pb	Amount of ^{210}Pb measured in the sample.
^{210}Pb error	Laboratory error associated with the ^{210}Pb measurement.
^{226}Ra	Amount of ^{226}Ra measured in the sample.
^{226}Ra	Laboratory error associated with the ^{226}Ra measurement.
Bulk sample: lab number	Laboratory number for the sample when submitted for 14C values. Sample numbers in the form of WW##### are from Lawrence Livermore, ones in the form of UCIT##### are from UC Irvine.
Bulk sample: fraction modern	Fraction modern for the bulk soil sample.
Bulk sample: \pm fraction modern	Analytical error associated with the fraction modern value for the bulk soil sample.
Bulk sample: $\Delta^{14}\text{C}$	$\Delta^{14}\text{C}$ value (‰) for the bulk soil sample.
Bulk sample: $\pm \Delta^{14}\text{C}$	Analytical error of $\Delta^{14}\text{C}$ value (‰) for the bulk soil sample.
Bulk sample: ^{14}C age	Radiocarbon age (years, uncorrected) of the bulk soil sample.
Bulk sample: $\pm ^{14}\text{C}$ age	Analytical error of radiocarbon age (years) for the bulk soil sample.
Picked material: lab number	Laboratory number for the sample when submitted for 14C values. Sample numbers in the form of WW##### are from Lawrence Livermore, ones in the form of UCIT##### are from UC Irvine.
Picked material: fraction modern	Fraction modern for picked material.
Picked material: \pm fraction modern	Analytical error of the fraction modern value for picked material.
Picked material: $\Delta^{14}\text{C}$	$\Delta^{14}\text{C}$ value for picked material.
Picked material: $\pm \Delta^{14}\text{C}$	Analytical error of $\Delta^{14}\text{C}$ value for picked material.
Picked material: ^{14}C age	Radiocarbon age (uncorrected) of picked material.
Picked material: $\pm ^{14}\text{C}$ age	Analytical error of radiocarbon age for picked material.
Notes	Information regarding what type of material was picked and submitted for ^{14}C analysis.

